Estimation of Antarctic land-fast sea ice algal biomass and snow thickness from under-ice radiance spectra in two contrasting 2 areas

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Key Points: 14

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15	•	New algorithms to estimate Antarctic land-fast sea ice algal biomass and snow
16		thickness from under-ice radiance spectra
17	•	The algorithms were developed for high biomass regimes in columnar and incorpo-
18		rated platelet ice
19	•	Inclusion of chlorophyll a-specific ice algal absorbance and pigment packaging
20		slightly affect biomass predictability

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21 Abstract

Fast ice is an important component of Antarctic coastal marine ecosystems, providing a 22 prolific habitat for ice algal communities. This work examines the relationships between 23 Normalized Difference Indices (NDI) calculated from under-ice radiance measurements 24 and sea ice algal biomass and snow thickness for Antarctic fast ice. While this technique 25 has been calibrated to assess biomass in Arctic fast ice and pack ice, as well as Antarctic 26 pack ice, relationships are currently lacking for Antarctic fast ice. We analyze measure-27 ments along transect lines at two contrasting Antarctic fast ice sites in terms of platelet 28 ice presence: near and distant from an ice shelf, i.e., in McMurdo Sound and off Davis 29 Station, respectively. Snow and ice thickness, and ice salinity and temperature measure-30 ments underpin our paired in situ optical and biological measurements. Analysis of our 31 samples shows that NDI wavelength pairs near the first chlorophyll-a (chl a) absorption 32 peak (≈ 440 nm) explain up to 70% of the total variability in algal biomass. Eighty eight 33 percent of snow thickness variability is explained using an NDI with a wavelength pair 34 of 648 nm and 567 nm. Accounting for pigment packaging effects by including the ratio 35 of chl a specific absorption coefficients improved the NDI-based algal biomass estimation 36 only slightly. Our new observation-based algorithms can be used to estimate Antarctic fast 37 ice algal biomass and snow thickness non-invasively, for example by using moored sensors 38 (time-series) or mapping their spatial distributions using underwater vehicles. 39

40 **1 Introduction**

Land-fast sea ice (fast ice) is an important feature of Antarctic coastal icescapes 41 [Fraser et al., 2012] and plays a key role in biogeochemical cycles and marine ecosystem 42 function [Arrigo, 2017]. Forming a thin barrier between the ocean and the atmosphere, 43 fast ice affects regional sea ice production [Tamura et al., 2016], forms a temporary reser-44 voir for nutrients [de Jong et al., 2013], and controls the amount of light available for local 45 pelagic primary production [Perovich, 2017]. Fast ice also serves as a habitat for micro-46 algal communities, which can contribute up to 50% to the overall primary production 47 of fast ice-covered areas [McMinn et al., 2010]. Fast ice algal communities are gener-48 ally concentrated in the lowermost portion of the ice, which is typically characterized by 49 the highly porous, 0.02-0.03 m thick skeletal layer [Weeks, 2010]. The bottom commu-50 nities are positioned in an optimal location to receive irradiance transmitted through the 51 ice, colonize an environment with low variability in temperature and salinity, and have ac-52 cess to replenishment of nutrients from the underlying seawater. Extremely high ice algal 53 biomass values have been reported for Antarctic fast ice containing platelet ice [Arrigo 54 et al., 1995; Günther and Dieckmann, 1999], which forms from supercooled Ice Shelf Wa-55 ter [Langhorne et al., 2015, and references therein] at the base of fast ice adjacent to ice 56 shelves [Morecki, 1965; Crocker and Wadhams, 1989]. This friable, highly porous sub-ice 57 platelet layer provides a large surface area that serves as an important algal micro-habitat. 58 Over time, it partially solidifies to form a bottom layer of incorporated platelet ice [e.g., 59 Gow et al., 1982; Smith et al., 2001]. 60

During early spring (October-November), Antarctic fast ice basal layers generally 61 provide a habitat for well over 90% of the total integrated fast ice algal biomass [Arrigo 62 et al., 1993; McMinn et al., 2010]. Although this is a low light environment, the algal 63 communities acclimate by modifying their photosynthetic apparatus through increasing 64 intracellular levels of chlorophyll-a (chl a) as well as accessory pigments, contributing to 65 so-called "pigment packaging" [Arrigo et al., 1993, 2014]. In self-shaded sea ice habi-66 tats where there is concentrated biomass, such as the sub-ice platelet layer [Arrigo et al., 67 1993], this adaptation enhances the algal light-capturing capacity by packaging more chl a68 per cell and simultaneously decreasing the absorption efficiency per unit pigment. Pigment 69 packaging is also affected by cell size [e.g., Duysens, 1956; Morel and Bricaud, 1981]. 70

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The high spatial variability of ice algal distribution has greatly hampered accurate 71 in situ estimation of ice algal biomass on relevant scales using classical sampling methods 72 such as ice coring which is both time consuming and limited in areal coverage [Mundy 73 et al., 2007; Lange et al., 2016; Meiners et al., 2017]. Ice coring is also destructive and 74 thus not well suited for time series measurements of ice algal development in a single 75 location [Campbell et al., 2014]. Here, we exploit a new method based on the effect of 76 ice algal photosynthetic pigments on the spectral composition of the under-ice light field 77 through absorption of specific wavelengths in the range of photosynthetically active radi-78 ation (PAR, 400 to 700 nm) [e.g., Perovich et al., 1993; Arrigo and Sullivan, 1994; Fritsen 79 et al., 2011; Hawes et al., 2012; Lange et al., 2016]. 80

A number of recent studies have explored the use of under-ice spectra to develop 81 algorithms to estimate sea ice algal chl a (as a proxy for biomass) from spectral under-82 ice irradiance or radiance data. Mundy et al. [2007] were the first to employ Normalized 83 Difference Indices (NDI) of under-ice light spectra to investigate the influence of both ice 84 algal biomass (i.e., chl a) and snow thickness on transmitted irradiance spectra under Arc-85 tic fast ice. Their study showed that snow has little effect on transmitted under-ice irradi-86 ances in the wavelength range of 400 to 570 nm and that a single NDI wavelength pair of 87 485:472 nm explained 89% of the ice algal variability observed in Arctic fast ice. Camp-88 bell et al. [2014] applied this NDI approach to monitor changes in basal chl a concentra-89 tion in Arctic fast ice. Their study highlighted the need for regional calibration of NDI 90 algorithms, i.e., regional adjustment for NDI to ice algal chl a conversions. Using an alter-91 native numerical method, Lange et al. [2016] recently described under ice irradiance-chl a 92 relationships for Arctic pack ice by an Empirical Orthogonal Function (EOF) approach. 93

For Antarctica, *Melbourne-Thomas et al.* [2015] provided regional NDI-to-sea ice chl *a* conversion algorithms for pack ice in two contrasting regions of the Southern Ocean, while *Forrest et al.* [2016] provided an initial algorithm for fast ice in McMurdo Sound (Ross Sea). Taken together, these studies suggest that NDI-chl *a* relationships vary regionally and cannot be applied universally as absorption of light might be influenced by characteristics other than total biomass and may, in particular, be influenced by variable ice algal pigment packaging.

¹⁰¹ By investigating relationships between sea ice physical properties, ice algal absorbance ¹⁰² spectra and under-ice irradiance spectra at similar times of the year, the aim of the present

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study is to develop and evaluate the first NDI-ice algal chl a algorithms for two contrast-103 ing Antarctic fast ice areas (Figure 1). Both are characterized by very high algal standing 104 stocks, i.e., up to an order of magnitude higher than observed in any of the previous NDI 105 studies. One, however, was in close proximity to the McMurdo Ice Shelf, Ross Sea, and 106 was characterized by the presence of platelet ice, while the other site off Davis Station, 107 East Antarctica, was influenced by neither an ice shelf nor platelet ice formation. Further-108 more, we provide a first assessment of the effects of pigment packaging on NDI-ice algal 109 biomass relationships. We also apply this technique to the estimation of snow thickness, 110 which is in itself an important parameter (both physical and biological) that requires more 111 extensive measurements [Sturm and Massom, 2017]. 112

113 2 Data and Methods

2.1 Study sites

A total of 67 paired under-ice radiance measurements and ice core samples were collected from 2 transects on Antarctic fast ice in McMurdo Sound (MS1 and MS2; N = 24) and 4 transects off Davis Station (DS1, DS2, DS3, and DS4; N = 43) in November 2014 and in November-December 2015, respectively (see Figure 1). Both the McMurdo Sound and Davis Station bio-optical sampling areas resembled undeformed first-year land-fast sea ice and were free of any surface disturbances.

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2.2 Integrated physical and biological measurements

122 In situ measurements

Figure 2 illustrates the integrated physical and biological measurement techniques 123 in McMurdo Sound (MS) and at Davis Station (DS). At each sampling site, we deployed 124 a TriOS Ramses ARC VIS radiometer (radiance sensor, 7° field of view) 0.15 m beneath 125 the bottom of the ice through small access holes (diameter 0.11 m and 0.25 m at DS and 126 MS, respectively) using a retractable L-shaped stainless steel arm (shown in Figure 2d). 127 The effect of the holes in contaminating light measurements was assumed to be negligible 128 due to the position of the radiometer 1.2 m north (i.e., directed towards the sun) of the 129 access hole [Melbourne-Thomas et al., 2015]. Under-ice high resolution radiance spectra 130 (320 to 950 nm, 3.3 nm bandwidth) were recorded with a laptop computer using TriOS 131 MSDA_XE software version 7.5.1. Results reported here represent average spectra from 132 three or more replicate spectral measurements. 133

Following the radiance measurements, snow thickness was measured with a ruler 134 (0.001 m gradation, 5 replicate measurements) at each site and one ice core was collected 135 directly above the radiometer location, using a Kovacs Mark II ice corer (0.09 m inter-136 nal diameter). The total length of the ice core was recorded and the 0.1 m bottom section 137 was cut off, placed in a clean polyethylene container and transported to the laboratory. 138 Sampled ice core sections were then melted at 4 °C in the dark, after adding 0.2 µm fil-139 tered seawater to avoid osmotic stress [Garrison and Buck, 1986]. After the ice cores had 140 melted (24 to 36 hours after collection), the samples were gently mixed and subsamples 141 were taken for a) determination of ice algal pigment concentration, and b) particulate ab-142 sorption. 143

In parallel with the biological measurements, two additional 0.09 m diameter cores 144 were extracted at 0 m and 128 m from each transect for standard sea ice temperature, salin-145 ity and structural measurements. One core from each site was used for thick section struc-146 tural analysis in a freezer laboratory. Selected thick sections were microtomed to produce 147 thin sections and were photographed. Temperature and salinity were measured on the 148 other cores, with the vertical temperature profiles being recorded immediately after ex-149 traction using an OMEGA Handheld Thermometer (OMEGA Engineering; Connecticut, 150 USA). Thereafter, the core was cut into 0.1 m sections which were placed in clean plastic 151 jars and transported to the laboratory for further analysis. Once melted at room tempera-152 ture, salinities were measured with a YSI 30 conductivity meter (YSI Inc.; Ohio, USA). 153

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Ice algal chlorophyll a concentration

For pigment analysis, 0.15 to 1.0 L melted ice core subsamples were filtered us-155 ing 25 mm diameter glass-fiber (Whatman GF/F) filters. The filters were then frozen and 156 stored below -80 °C for analysis using High Performance Liquid Chromatography (HPLC). 157 Samples were extracted over 15 to 18 hours in acetone before analysis by HPLC using 158 a modified C8 column and binary gradient system with an elevated column temperature 159 [Clementson, 2013]. Chl a was identified by retention time and absorption spectrum from 160 a photo-diode array detector, and concentrations were determined from commercial and 161 international standards (Sigma; DHI, Denmark). 162

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Particulate (algal and non-algal) absorption

For absorption analyses, melted ice core subsamples (0.15 to 1.0 L) were filtered 164 onto 25 mm diameter glass-fiber (Whatman GF/F) filters. The optical density (OD) spectra 165 of the particulate material on these filters were measured over the 350 to 750 nm spec-166 tral range in 0.9 nm increments, using a Cintra 404 UV/VIS dual-beam spectrophotome-167 ter equipped with an integrating sphere. The pigments on the sample filter were then ex-168 tracted following the method of Kishino et al. [1985] to determine the OD of the non-algal 169 particles in a second scan. The OD due to ice algae was derived by calculating the dif-170 ference between the optical density of the total particulate and non-algal fractions. The 171 OD measurements were converted to absorption spectra using blank filter measurements, 172 and by first normalizing the scans to zero at 750 nm and then correcting for the path 173 length amplification using the coefficients of *Mitchell* [1990]. A detailed description of the 174 method is given in Clementson et al. [2001], following ocean-optics protocols for SeaWiFS 175 validation [Müller et al., 2003]. 176

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based on:

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$$a_{\text{nap}}(\lambda) = a_{\text{nap}}(350\,\text{nm})\exp[-S(\lambda - 350\,\text{nm})] + b,\tag{1}$$

where $a_{nap}(\lambda)$ is the residual absorption coefficient over the wavelength (λ) range 350 to 750 nm 180 of the particles after methanol extraction, also referred to as absorption of non-algal parti-181 cles $[m^{-1}]$ which includes absorption of non-extractable pigments and heterotrophic pro-182 tists. A non-linear least-squares technique was used to fit Equation 1 to the untransformed 183 data, where S and b are empirically-determined constants. The inclusion of offset b al-184 lows for any baseline correction. In some samples, pigment extraction was incomplete, 185 leaving small residual peaks in the non-algal particulate spectra at the principal chloro-186 phyll absorption bands. To avoid distorting the fitted non-algal particulate spectra, data 187 at these wavelengths were omitted when all spectra were fitted. Total particulate spectra 188 were smoothed using a running box-car filter with 10 nm width, and the fitted non-algal 189 particulate spectra were subtracted to yield the ice algae spectra. 190

An exponential function was fitted to all spectra of non-algal particulate material

The following parameters were then determined: $a_{\rm p}(\lambda)$ = absorption coefficient of particles [m⁻¹]; $a_{\rm ph}(\lambda)$ = absorption coefficient of ice algae [m⁻¹] calculated as the difference between $a_{\rm p}(\lambda)$ and $a_{\rm nap}(\lambda)$; and the chl *a* specific absorption coefficient of ice algae $a_{\rm ph}^*(\lambda)$ [m²(mg chl *a*)⁻¹] which is equal to the ratio of $a_{\rm ph}(\lambda)$ and (chl *a*) concentration.

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¹⁹⁵ Note that the resulting ice algae spectra were base-corrected by subtracting absorption at

¹⁹⁶ 750 nm to obtain $a_{\rm ph}(\lambda)$. We used non-parametric Pearson's correlation and linear regres-

¹⁹⁷ sion analyses to explore the relationships between variables.

198 **2.3 Spectral Data Analyses**

199 Normalized Difference Index

All transmitted radiance spectra were interpolated to consecutive integer (1 nm) wavelengths and restricted to the range from 400 to 700 nm. To visualize correlations between integrated chl *a* and under-ice transmitted radiance (I_T) measurements, we followed the approach of *Mundy et al.* [2007]. We first calculated NDI for all possible wavelength pairs according to the formula:

NDI
$$(\lambda_1, \lambda_2) = \frac{I_T(\lambda_1) - I_T(\lambda_2)}{I_T(\lambda_1) + I_T(\lambda_2)},$$
 (2)

where λ_1 and λ_2 are wavelengths in the PAR range 400 to 700 nm. We then correlated the derived NDI values with integrated chl *a* concentrations and snow thickness data, and plotted the resultant Pearson's correlation coefficient surfaces.

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Best NDI (λ_1, λ_2) selection

Following Mundy et al. [2007], spectral surfaces of Pearson's correlation coefficients 210 between NDI of under-ice transmitted radiance from all possible wavelength combina-211 tions and $log_{10}(chl a)$ and snow thickness were constructed for groups MS, DS and com-212 bined MS and DS. Note that in Mundy et al. [2007], NDI is a response variable and chl a 213 a predictor variable. Subsequently, Campbell et al. [2014] swapped chl a to response to re-214 flect the applicability of using NDI as a predictor. Natural logarithm was applied to chl a 215 (ln(chl a)) to deal with the high variance at high chl a (heteroscedasticity) in Melbourne-216 Thomas et al. [2015] and for model comparison purposes in Lange et al. [2016]. In this 217 study the range of our chl a dataset covers three orders of magnitude so we follow previ-218 ous studies but replace $\ln(\operatorname{chl} a)$ with $\log_{10}(\operatorname{chl} a)$. 219

To derive an optimal NDI wavelength pair (λ_1, λ_2) for both chl *a* and snow, we applied the following inputs. Firstly and in addition to the Pearson's correlation surfaces for chl *a* and snow, we constructed coefficient of determination (R^2) surfaces by using $\log_{10}(\text{chl } a)$ or snow thickness as a response and NDI (λ_1, λ_2) as a predictor for all wave-

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length pairs. Secondly, we only considered NDI consisting of wavebands λ_1 and λ_2 that 224 are at least 15 nm (> 3 radiometer channels) apart to ensure clear separation when de-225 tected by the RAMSES radiometer. For chl a, wavelengths between 405 nm and 550 nm 226 avoid both edge effects and the influence of snow on transmitted radiance spectra beyond 227 550 nm. For snow, the optimal wavelength pairs are between 405 nm and 655 nm to avoid 228 proximity to the second strong absorption peak of chl a at ≈ 670 nm [e.g., Mundy et al., 229 2007]. In addition, the null hypothesis that $\log_{10}(\operatorname{chl} a)$ is constant (i.e., does not depend 230 on NDI(λ_1, λ_2)) was tested against the alternative hypothesis that $\log_{10}(\text{chl } a)$ depends 231 linearly on NDI(λ_1 , λ_2). If the p-value is less than a critical value (set to 0.05), then the 232 null hypothesis is rejected at the 5% significance level. 233

Pigment packaging index

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²³⁵ We sought to improve the inter-relationships among NDI and chl *a* by considering ²³⁶ the effect of pigment packaging. The chl *a* specific absorption coefficients $(a_{ph}^*(\lambda)$ from ²³⁷ absorption peaks centered at ≈ 440 and ≈ 676 nm) were used to construct a Pigment Pack-²³⁸ aging Index (PPI) [*Arrigo et al.*, 2014] as:

$$PPI = \frac{a_{ph}^{*}(440 \text{ nm})}{a_{ph}^{*}(676 \text{ nm})}.$$
(3)

The inter-relationship of NDI-chl *a*-PPI was explored by using the following procedure. After selecting the best pairs of NDI for chl *a*, $\log_{10}(PPI)$ was added as another predictor. The null hypothesis that $\log_{10}(chl a)$ depends linearly on NDI(λ_1 , λ_2), was tested against the alternative hypothesis that $\log_{10}(chl a)$ depends on NDI(λ_1 , λ_2) and $\log_{10}(PPI)$.

To compare models with different number of predictors, i.e., the chl *a* model with and without PPI, the adjusted R^2 (R^2_{adj}) was used instead of R^2 as a measure of goodness of fit. R^2 continues to increase with more predictors, while adjusts for the number of predictors [e.g., *Chatterjee and Hadi*, 2012]. Thus if the difference between R^2 and R^2_{adj} is small, then the model(s) do not overfit the data. 249 **3 Results**

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3.1 Physical property measurements

We first summarize the snow and ice thickness, ice salinity and ice temperature measurements that underpin our in situ optical and biological paired measurements. Snow cover thickness was small and less variable at McMurdo Sound (mean \pm 1 standard deviation (sd, hereafter): 0.01 \pm 0.02 m; range: 0 to 0.11 m) compared to Davis Station (mean \pm sd: 0.27 \pm 0.09 m m; range: 0.09 to 0.47 m). At the former, zero snow thickness due to wind scouring occurred locations at 0.5 and 1 m on transect MS1 and 16 m on MS2.

Figure 3 shows plots of all temperature and salinity profiles and representative basal 257 sea ice structures from both sampling areas. The fast ice at McMurdo Sound (MS) was 258 thicker than that at Davis Station (DS). Ice thickness at MS sampling sites ranged be-259 tween 1.91 and 2.16 m (mean \pm sd: 2.03 \pm 0.10 m, Figures 3a-b) and the sea ice was pre-260 dominantly columnar first-year, with an incorporated platelet ice layer at the bottom. A 261 very thin (< 0.02 m) sub-ice platelet layer was observed at the base of the sea ice at MS 262 and the transition from columnar to incorporated platelet ice occurred at 1.75 (1.90) m 263 and 1.78 (1.51) m for the 0 (512) m cores at MS1 and MS2, respectively. For DS sites, 264 on the other hand, columnar sea ice was present through most of the thickness (mean \pm 265 sd:1.55 \pm 0.06 m; range: 1.48 to 1.64 m), with a thin (~ 0.1 m) layer of snow ice at the 266 upper interface. Thus the bottom 0.1 m sections of sea ice cores from MS were purely 267 incorporated platelet ice (Figure 3d) while those for DS were purely columnar (Figure 3f). 268

Ice temperature profiles with depth were approximately linear during both sampling 269 campaigns, ranging from -9.2 to -1.6 °C at MS and -5.1 to -1.6 °C at DS (Figure 3a). 270 Diurnal temperature deviations from linearity were observed primarily near the sea ice 271 surface due to the near-surface air temperature variability. Temperatures near the sea ice 272 bottom were similar and close to the freezing temperature of seawater. Sea ice salinities 273 ranged from 3.7 to 13.4 ppt and from 2.9 to 15.2 ppt at MS and DS, respectively (Fig-274 ure 3b). Salinity profiles showed a typical C-shape with comparatively high values at the 275 top and (particularly) the bottom of the cores. 276

3.2 Under-ice radiance hyperspectral measurements and chl a

Mean \pm sd under-ice radiance spectra normalized by the area under the curves in the 400 to 700 nm range for McMurdo Sound (MS; N = 24) and Davis Station (DS; N = 43) are shown in Figure 4a. Both MS and DS normalized spectra show local minima at around 440 and 675 nm at known chl *a* absorption peaks. Higher ice algal biomass reduces transmitted radiance in the blue part of the spectrum and results in a compressed peak in the green part of the spectrum [*Legendre and Gosselin*, 1991].

In McMurdo Sound, chl *a* concentrations in the lowermost 0.1 m of the cores varied between 285.4 and 2109.5 mg m⁻³ (mean \pm sd: 1132.5 \pm 414.8 mg m⁻³). At Davis Station, concentrations were lower with a range of 2.0 to 1721.0 mg m⁻³ (mean \pm sd: 580.5 \pm 441.8 mg m⁻³). The corresponding ice algal chlorophyll *a* contents (integrated over the lowermost 0.1 m of the ice) ranged from 26.0 to 172.4 mg m⁻² (mean \pm sd: 105.2 \pm 36.3 mg m⁻²) and from 0.2 to 165.0 mg m⁻² (mean \pm sd: 55.1 \pm 42.4 mg m⁻²) at MS and DS, respectively (Figure 4b).

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3.3 Particulate absorption measurements and pigment packaging index

The mean and standard deviation of the absorption spectra are plotted in Figure 5 292 for both sites. Absorption spectra of all particulate matter $(a_{\rm p})$ demonstrate strong ab-293 sorption by chl a with distinct maxima at 440 and 675 nm (Figure 5a). Non-algal partic-294 ulate matter spectra (a_{nap}) , as shown in Figure 5b, decrease exponentially with increasing 295 wavelength. The ice algal absorption spectra (a_{ph}) and the chl *a* specific absorption coef-296 ficients (Figures 5c and d, respectively) follow the form of a_p . However, the mean of MS 297 is smaller than that of DS due to the higher average chl a concentrations at MS site (Fig-298 ure 5d). 299

The dependence of chl *a* and PPI on snow thickness is shown in Figure 6a-b. As defined in Equation 3, PPI is constructed from as the ratio of the two chl *a* absorption maxima. Values of PPI varied from 1.25 to 2.09 with a mean of 1.67 ± 0.16 (Figure 6b). Both chl *a* and PPI are negatively correlated with snow depth, with Pearson's correlation coefficient values (ρ) of -0.69 and -0.45, respectively. The decrease in PPI indicates a flattening of the chl *a* specific absorption spectrum, i.e., increased pigment packaging.

306 3.4 NDI algorithm

Pearson's correlation coefficient surfaces (Figure 7) as well as R^2 surfaces (Figure 8) 307 were constructed for both chl a and snow thickness. The strong negative relationship be-308 tween biomass and snow thickness (Figure 6a) causes the filled contours of chl a in Fig-309 ures 7a, c, and e to be approximately the inverse of snow thickness in Figures 7b, d and 310 f. The R^2 surfaces of Figure 8 are used as the main criteria for selecting the optimal 311 wavelength pairs for the algorithm. Values of R^2 , R^2_{adj} and p-value for MS, DS and com-312 bined MS and DS are summarized in Table 1. In particular, optimal values of NDI(λ_1, λ_2) 313 of combined MS and DS for chl a and snow thickness are plotted in Figure 8. The best 314 wavelength pairs are separated by 15 nm and 81 nm for chl a and snow thickness, respec-315 tively. The root mean squared errors of data about the best fit model are 0.4 for $\log_{10}(\text{chl }a)$ 316 and 0.05 m for snow thickness. 317

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3.5 NDI-PPI-chl *a* interrelationships

To investigate the influence of the algal pigment packaging effect on NDI-based al-319 gal biomass estimates, R^2 , R^2_{adj} and p-values of a model of MS, DS, and combined MS 320 and DS were compared with a model that included PPI as a second predictor. The results 321 are summarized in Table 2. The relations for combined MS and DS data are shown in 322 Figure 9. The inclusion of the second predictor $\log_{10}(PPI)$ increases from 0.69 to 0.74. 323 Specifically, the significance of the log_{10} (PPI) term for combined MS and DS chl *a* was 324 tested with the null hypothesis that the description of chl a without PPI is adequate. The 325 p-value is 0.0005 (< 0.05), rejecting the null hypothesis. 326

327 4 Discussion

Our study provides empirical algorithms to estimate sea ice algal chl *a* content and snow thickness from under-ice radiance measurements at two physically different Antarctic fast ice locations that were both characterized by high ice algal biomass standing stocks.

The physical contrast between the MS and DS sites is mainly due to the difference in snow thickness and the presence of an incorporated platelet ice layer at MS. The sea ice at MS was thicker than at DS, partly because the growth rate of incorporated platelet ice is greater than that of purely columnar ice [*Smith et al.*, 2015]. Salinity measurements from both sites agreed with observations reporting C-shape profiles as typical for Antarctic first-year fast ice [e.g., *Eicken*, 1992; *Weeks*, 2010]. Salinities at MS corresponded with
 those reported by *Gough et al.* [2012], demonstrating only marginal differences between
 columnar and incorporated platelet ice.

Our sea ice structural analyses suggest that the basal ice algae inhabited different ice 339 types: incorporated platelet ice at MS and columnar ice at DS. Considering both columnar 340 (DS) and incorporated platelet ice (MS) as congelation ice with basal skeletal layers, sea 341 ice algae from both sites can be classified as bottom communities. The mean integrated 342 chl a concentrations at MS were higher than that at DS. Ice algal biomass standing stocks 343 are in agreement with previous studies conducted during the spring ice algal bloom period 344 at these sites [Arrigo et al., 1995; Archer et al., 1996; Arrigo, 2017]. The friable sub-ice 345 platelet layer that preceded the incorporated platelet ice at the base of the sea ice cover 346 in McMurdo Sound provided a porous habitat for high chl a accumulation [Arrigo et al., 347 1995; Günther and Dieckmann, 1999]. 348

Normalized under-ice radiance spectra show local minima at around 440 nm and 349 676 nm at all stations, consistent with the in situ absorbance maxima of ice algae [e.g., 350 Beeler SooHoo et al., 1987; Fritsen et al., 2011]. Our observations confirm previous re-351 ports on the influence of ice algal biomass on under-ice spectra from both the Arctic and 352 Antarctic [Beeler SooHoo et al., 1987; Perovich et al., 1993; Mundy et al., 2007; Ehn et al., 353 2008]. The chl *a* specific absorption coefficients $a_{ph}^*(\lambda)$ resemble the shape of the $a_p(\lambda)$ 354 spectra with maximum absorption in the blue-green region of the spectrum between 430 nm 355 and 530 nm, with a second narrower peak at around 676 nm (Figure 5d). An exponential 356 decrease of $a_{nap}(\lambda)$, the non-algal particle absorption coefficient, with increasing wave-357 length is typical for marine detrital material [Kishino et al., 1985, Figure 5b]. Variations 358 in $a_{ph}^*(\lambda)$ have been attributed to the algal community taxonomic composition, with cou-359 pled changes in intracellular pigment concentrations and composition as well as cell size 360 being interpreted as the pigment packaging effect [Arrigo et al., 1993; Bricaud, 2004]. 361 Spectra of $a_{\rm ph}^*(\lambda)$ observed in the present study correspond to the absorption of diatoms 362 containing chl a and accessory photosynthetic carotenoid pigments, particularly fucox-363 anthin which has maximum absorption around 450 to 550 nm [e.g., Arrigo et al., 1991]. 364 Diatoms generally dominate Antarctic fast ice bottom ice algal communities [Arrigo, 2014, 365

366 2017].

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Our empirical relationships from MS and DS and the combined dataset are compared with results from previous NDI studies in Table 3. The present study fills an NDI algorithm gap for Antarctic land-fast sea ice and also provides new algorithms that are calibrated for high chl *a* regimes in columnar and platelet ice (\approx 73 mg m⁻², range: 0.2 to 172.4 mg m⁻²) and snow depths ranging between 0 to 0.5 m (\approx 0.2 m).

Considering all algorithms in Table 3 suggests that if the snow is thick enough (> 372 0.1 m), the model selection method favors an NDI wavelength pair (for chl *a* estimation) 373 that is closest to the first chl a absorption peak (≈ 440 nm). When the snow is thin, then 374 model selection picks the highest correlation between NDI and chl a in the region of the 375 second peak (≈ 670 nm). Both NDI algorithms of *Lange et al.* [2016] are in this category, 376 with snow thickness less than 0.1 m. This can be interpreted as the influence of snow on 377 NDI-based chl a estimates at these higher wavelengths, as previously discussed by Mundy 378 et al. [2007]. To avoid interference of the second chl a absorption peak on the snow algo-379 rithm, we limited the search for the best NDI-snow algorithm to be in the 405 to 655 nm 380 (Table 1). From inspection, the best NDI for snow is not close to the first chl a absorption 381 peak, so we did not constrain the lower bound to avoid its interference with the algorithm. 382

As stated by *Mundy et al.* [2007] and illustrated in our R^2 surfaces, the NDI wavelength range between 400 to 550 nm is not well correlated with snow depth. Our NDI(441,426) correlates well with chl *a* ($\rho = -0.83$) and less with snow ($\rho = 0.70$) (Figure 8). Compared to DS, the optimal wavelength pair of MS was positioned at higher wavelengths than the first chl *a* absorption peak (> 440 nm). This is consistent with the findings of *Beeler SooHoo et al.* [1987], who reported enhanced absorption of platelet ice algae in the 450 to 550 nm wavelength range.

Our mean PPI (1.67 \pm 0.16) agrees with PPI values (2.02 \pm 0.46) reported for bottom 390 ice algal communities in Amundsen Sea pack ice [Arrigo et al., 2014]. The PPI model's 391 difference in R^2 and R^2_{adj} is small and demonstrates that the model does not overfit the 392 data by adding $log_{10}(PPI)$ as the second predictor after NDI. The p-value for $log_{10}(PPI)$ 393 term is 0.0005 (< 0.05), implying that this term is significant at the 5% significance level 394 for combined MS and DS. For the combined MS and DS dataset, inclusion of the pre-395 dictor $\log_{10}(\text{PPI})$ improves the R_{adi}^2 for chl *a* by up to 0.05 (see Tables 1 and 2). Given 396 this small improvement and the considerable effort needed for ice algae absorption mea-397 surements, we suggest that the inclusion of PPI can possibly be neglected for fast ice. 398

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However, the inclusion of PPI may become important when constructing integrated multiparameter algorithms involving different sea ice types (e.g., pack ice and fast ice) that
have higher variability of PPI due to different sea ice algal habitats [e.g., *Arrigo et al.*,
2014]. Recently, *Kauko et al.* [2017] also suggested that in Arctic leads, both light and
algal species composition influence pigment packaging.

Another way to improve the performance of optical algorithms to estimate ice algal 404 biomass was recently proposed by Lange et al. [2016] who suggested the application of 405 the EOF approach over NDI and proposed using transmittance data for algorithm devel-406 opment based on [Nicolaus and Katlein, 2013]. Our R^2 values from NDI algorithms are 407 similar to those reported in the study of Lange et al. [2016] (73% for NDI-irradiance and 408 70% for NDI-transmittance). However, the wavelengths selected for the NDI algorithms 409 of Lange et al. [2016] are close to the second chl a absorption peak at 670 nm which cor-410 relates with snow thickness [e.g., Mundy et al., 2007]. Note that the NDI algorithms of 411 Lange et al. [2016] were calibrated with both thin snow thickness data (< 0.1 m) and low 412 biomass. This might limit their applicability to Arctic sea ice with a thin snow cover, e.g., 413 late summer conditions when most of their data were collected. In Antarctic pack ice, the 414 NDI algorithms in Melbourne-Thomas et al. [2015] were optimized for low chl a concen-415 trations and thicker snow. Compared with other algorithms, including the EOF, they show 416 relatively high R^2 values for the NDI method. Lange et al. [2016] also suggested normal-417 izing spectra to their corresponding downwelling irradiance when dealing with datasets 418 taken over larger areas and during different seasons. This is particularly true for Arctic 419 sea ice that shows strong seasonal changes in surface conditions, i.e., changing from snow 420 cover to white ice and melt ponds before breaking up. 421

A relatively poor NDI-to-chl a relationship was observed for the MS study site. We 422 attribute this to: i) the low overall variability in algal biomass in this particular dataset; 423 and ii) potential biases in sampling the fragile unconsolidated sub-ice platelet layer at the 424 bottom of MS ice cores. Further work is required to advance quantitatively robust sam-425 pling techniques for platelet ice and to develop optical methods to understand the phe-426 nology of platelet ice algal communities as well as their spatial variability [Forrest et al., 427 2016]. Many of the MS sites also showed zero to very low snow depths but variable chl a428 content, suggesting that factors other than snow control ice algal biomass accumulation 429 in this area. The variability most likely relates to processes within the sub-ice platelet 430 layer habitat (Figure 2c) in which crystal structure and surface area, as well as nutrient 431

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availability within the porous ice matrix, are important for algal colonization [e.g., *Arrigo*,
2017].

Our results show that chl a was negatively correlated with snow depth. Strong in-434 verse relationships between snow cover thickness and ice algal standing stock have been 435 reported elsewhere for Antarctic sea ice habitats [Grossi and Sullivan, 1985; Ackley and 436 Sullivan, 1994]. For Arctic fast ice, a seasonally-changing influence of snow on ice al-437 gal standing stocks has been described [Mundy et al., 2005; Leu et al., 2015]. There, snow 438 limits light availability early in the ice-growth season, but the relationship reverses later 439 in the season, i.e., in boreal spring, because the snow cover prevents basal ice ablation, 440 thereby stabilizing the ice algal habitat [Campbell et al., 2015]. Thereafter formation of 441 melt ponds during summer allows higher light transmittance into Arctic sea ice [e.g., 442 Nicolaus et al., 2012]. Surface melt pond formation is rare on Antarctic sea ice [Andreas 443 and Ackley, 1982; Sturm and Massom, 2017]. A comparative description of ice algal phe-444 nology is lacking for Antarctic fast ice and future research is needed to investigate a po-445 tentially seasonally-changing snow-chl a relationship in Antarctic sea ice [Meiners et al., 446 2017]. 447

448 5 Conclusion

In conclusion, our study provides a first bio-optical investigation of first-year fast 449 ice from two contrasting areas of the Antarctic coast. The contrasts between the two sites 450 were mainly due to a higher variation in snow thickness at Davis Station sites, and the 451 presence of the incorporated platelet ice at McMurdo Sound sites due to the influence of 452 the McMurdo Ice Shelf. The NDI algorithm developed explains 70% of chl *a* for com-453 bined MS and DS data with wavelength pairs that are close to the first absorption peak 454 of chl a at around 440 nm. This result is consistent with observations in earlier studies 455 and extends the NDI algorithm towards a high chl a regime. Furthermore, NDI-based pre-456 dictability of algal biomass can be improved by up to 5% when the ratio of chl a specific 457 absorption of both peaks is included in the model. 458

Another important finding was that the NDI developed from the wavelength combination above 550 nm and below the second chl *a* absorption peak describes 88% of the variation in the combined MS and DS snow thickness data. Our empirical relationship for the MS site was limited, however, and future work is required to develop robust relation-

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ships for sea ice with a significant platelet ice component. The interrelationships of chl a, 463 snow and NDI provide an important step towards developing a calibrated toolbox to ob-464 tain chl a and potentially snow depth at local to regional scales using remotely-operated or 465 autonomous underwater vehicles. Deployment of hyperspectral under-ice radiometers pro-466 vides a non-invasive tool for ice algal biomass determination. Such radiometers should be 467 routinely incorporated into physical sea ice mass-balance stations collecting data through-468 out the sea-ice season to further our understanding of coupled sea ice physical-biological 469 processes and ice algal phenologies in polar ecosystems. 470

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Data availability: All data used in this study are available through the Australian 487 Antarctic Data Centre (AADC) at https://data.aad.gov.au. 488

-17-

489 **References**

- 490 Ackley, S., and C. Sullivan (1994), Physical controls on the development and character-
- 491 istics of Antarctic sea ice biological communities— a review and synthesis, *Deep-Sea*
- Research Part I: Oceanographic Research Papers, 41(10), 1583–1604, doi:10.1016/0967 0637(94)90062-0.
- Andreas, E. L., and S. F. Ackley (1982), On the differences in ablation seasons of Arctic and Antarctic sea ice, *Journal of the Atmospheric Sciences*, *39*(2), 440–447, doi:
- ⁴⁹⁶ 10.1175/1520-0469(1982)039<0440:otdias>2.0.co;2.
- ⁴⁹⁷ Archer, S., R. Leakey, P. Burkill, M. Sleigh, and C. Appleby (1996), Microbial ecology
- of sea ice at a coastal Antarctic site: community composition, biomass and temporal
 change, *Marine Ecology Progress Series*, *135*, 179–195, doi:10.3354/meps135179.
- Arrigo, K. R. (2014), Sea ice ecosystems, *Annual Review of Marine Science*, 6(1), 439–
 467, doi:10.1146/annurev-marine-010213-135103.
- Arrigo, K. R. (2017), Sea ice as a habitat for primary producers, in Sea
- Ice, edited by D. N. Thomas, pp. 352–369, John Wiley & Sons, Ltd, doi:
- ⁵⁰⁴ 10.1002/9781118778371.ch14.
- Arrigo, K. R., and C. W. Sullivan (1994), A high resolution bio-optical model of mi croalgal growth: Tests using sea-ice algal community time-series data, *Limnology and Oceanography*, *39*(3), 609–631, doi:10.4319/lo.1994.39.3.0609.
- Arrigo, K. R., C. W. Sullivan, and J. N. Kremer (1991), A bio-optical model of Antarctic sea ice, *Journal of Geophysical Research*, *96*(C6), 10,581, doi:10.1029/91jc00455.
- Arrigo, K. R., D. H. Robinson, and C. W. Sullivan (1993), High resolution study of the
- ⁵¹¹ platelet ice ecosystem in McMurdo Sound, Antarctica: photosynthetic and biooptical
- characteristics of a dense microalgal bloom, *Marine Ecology Progress Series*, 98, 173–
 185, doi:10.3354/meps098173.
- Arrigo, K. R., G. Dieckmann, M. Gosselin, D. H. Robinson, C. H. Fritsen, and C. W. Sul-
- livan (1995), High resolution study of the platelet ice ecosystem in McMurdo Sound,
- Antarctica: biomass, nutrient, and production profiles within a dense microalgal bloom,
- 517 *Marine Ecology Progress Series*, 127, 255–268, doi:10.3354/meps127255.
- Arrigo, K. R., Z. W. Brown, and M. M. Mills (2014), Sea ice algal biomass and phys-
- ⁵¹⁹ iology in the Amundsen Sea, Antarctica, *Elementa: Science of the Anthropocene*, 2,
- ⁵²⁰ 000,028, doi:10.12952/journal.elementa.000028.

521	Beeler SooHoo, J., A. C. Palmisano, S. T. Kottmeier, M. P. Lizotte, S. L. SooHoo, and
522	C. W. Sullivan (1987), Spectral light absorption and quantum yield of photosynthesis in
523	sea ice microalgae and a bloom of Phaeocystis pouchetii from Mcmurdo Sound, Antarc-
524	tica, Marine Ecology Progress Series, 39, 175–189, doi:10.3354/meps039175.
525	Bricaud, A. (2004), Natural variability of phytoplanktonic absorption in oceanic waters:
526	Influence of the size structure of algal populations, Journal of Geophysical Research,
527	109(C11), doi:10.1029/2004jc002419.
528	Campbell, K., C. J. Mundy, D. G. Barber, and M. Gosselin (2014), Remote estimates of
529	ice algae biomass and their response to environmental conditions during spring melt,
530	ARCTIC, 67(3), 375, doi:10.14430/arctic4409.
531	Campbell, K., C. Mundy, D. Barber, and M. Gosselin (2015), Characterizing the sea ice
532	algae chlorophyll a-snow depth relationship over Arctic spring melt using transmitted
533	irradiance, Journal of Marine Systems, 147, 76-84, doi:10.1016/j.jmarsys.2014.01.008.
534	Chatterjee, S., and A. S. Hadi (2012), Regression Analysis by Example, John Wiley &
535	Sons, Inc.
536	Clementson, L. A. (2013), The CSIRO method, in The Fifth SeaWiFS HPLC Analysis
537	Round-Robin Experiment (SeaHARRE-5), edited by S. B. Hooker, L. A. Clementson,
538	C. S. Thomas, L. Schlüter, M. Allerup, J. Ras, H. Claustre, C. Normandeau, J. Cullen,
539	M. Kienast, W. Kozlowski, M. Vernet, S. Chakraborty, S. Lohrenz, M. Tuel, D. Redalje,
540	P. Cartaxana, C. R. Mendes, V. Brotas, S. G. P. Matondkar, S. G. Parab, A. Neeley, and
541	E. S. Egeland, NASA Technical Memorandum 2012-217503, NASA Goddard Space
542	Flight Center, Greenbelt, Maryland.
543	Clementson, L. A., J. S. Parslow, A. R. Turnbull, D. C. McKenzie, and C. E. Rath-
544	bone (2001), Optical properties of waters in the Australasian sector of the South-
545	ern Ocean, Journal of Geophysical Research: Oceans, 106(C12), 31,611-31,625, doi:
546	10.1029/2000jc000359.
547	Crocker, G., and P. Wadhams (1989), Modelling Antarctic fast-ice growth, Journal of
548	Glaciology, 35(119), 3-8, doi:10.3189/002214389793701590.
549	de Jong, J., V. Schoemann, N. Maricq, N. Mattielli, P. Langhorne, T. Haskell, and JL.
550	Tison (2013), Iron in land-fast sea ice of McMurdo Sound derived from sediment resus-
551	pension and wind-blown dust attributes to primary productivity in the Ross Sea, Antarc-
552	tica, Marine Chemistry, 157, 24-40, doi:10.1016/j.marchem.2013.07.001.

-19-

- Duysens, L. (1956), The flattening of the absorption spectrum of suspensions, as com-553
- pared to that of solutions, Biochimica et Biophysica Acta, 19, 1-12, doi:10.1016/0006-554 3002(56)90380-8. 555
- Ehn, J. K., C. J. Mundy, and D. G. Barber (2008), Bio-optical and structural properties in-556 ferred from irradiance measurements within the bottommost layers in an Arctic landfast 557
- sea ice cover, Journal of Geophysical Research, 113(C3), doi:10.1029/2007jc004194. 558
- Eicken, H. (1992), Salinity profiles of Antarctic sea ice: Field data and model results, 559 Journal of Geophysical Research, 97(C10), 15,545, doi:10.1029/92jc01588.
- and I. Hawes (2016), Brief communication: Capturing scales of spatial heterogeneity 562 of Antarctic sea ice algae communities, The Cryosphere Discussions, pp. 1-13, doi: 563

Forrest, A. L., L. C. Lund-Hansen, B. K. Sorrell, I. Bowden-Floyd, V. Lucieer, R. Cossu,

10.5194/tc-2016-186. 564

560

561

- Fraser, A. D., R. A. Massom, K. J. Michael, B. K. Galton-Fenzi, and J. L. Lieser (2012), 565
- East Antarctic landfast sea ice distribution and variability, 2000-08, Journal of Climate, 25(4), 1137-1156, doi:10.1175/jcli-d-10-05032.1. 567
- Fritsen, C. H., E. D. Wirthlin, D. K. Momberg, M. J. Lewis, and S. F. Ackley (2011), 568
- Bio-optical properties of Antarctic pack ice in the early austral spring, Deep Sea 569
- Research Part II: Topical Studies in Oceanography, 58(9-10), 1052–1061, doi: 570
- 10.1016/j.dsr2.2010.10.028. 571
- Garrison, D. L., and K. R. Buck (1986), Organism losses during ice melting: A 572 serious bias in sea ice community studies, Polar Biology, 6(4), 237-239, doi: 573 10.1007/bf00443401. 574
- Gough, A. J., A. R. Mahoney, P. J. Langhorne, M. J. M. Williams, and T. G. Haskell 575 (2012), Sea ice salinity and structure: A winter time series of salinity and its distribu-576
- tion, Journal of Geophysical Research: Oceans, 117(C3), doi:10.1029/2011jc007527. 577
- Gow, A., S. Ackley, W. Weeks, and J. Govoni (1982), Physical and structural 578
- characteristics of Antarctic sea ice, Annals of Glaciology, 3, 113-117, doi: 579
- 10.1017/s0260305500002627. 580
- Grossi, S. M., and C. W. Sullivan (1985), Sea ice microbial communities. V. the vertical 581 zonation of diatoms in an Antarctic fast ice community, Journal of Phycology, 21(3), 582
- 401–409, doi:10.1111/j.0022-3646.1985.00401.x. 583
- Günther, S., and G. S. Dieckmann (1999), Seasonal development of algal biomass in 584
- snow-covered fast ice and the underlying platelet layer in the Weddell Sea, Antarctica, 585

586	Antarctic Science, 11(03), doi:10.1017/s0954102099000395.
587	Hawes, I., L. C. Lund-Hansen, B. K. Sorrell, M. H. Nielsen, R. Borzák, and I. Buss
588	(2012), Photobiology of sea ice algae during initial spring growth in Kangerlussuaq,
589	West Greenland: insights from imaging variable chlorophyll fluorescence of ice cores,
590	Photosynthesis Research, 112(2), 103-115, doi:10.1007/s11120-012-9736-7.
591	Kauko, H. M., T. Taskjelle, P. Assmy, A. K. Pavlov, C. J. Mundy, P. Duarte,
592	M. Fernández-Méndez, L. M. Olsen, S. R. Hudson, G. Johnsen, A. Elliott, F. Wang,
593	and M. A. Granskog (2017), Windows in Arctic sea ice: Light transmission and ice al-
594	gae in a refrozen lead, Journal of Geophysical Research: Biogeosciences, 122(6), 1486-
595	1505, doi:10.1002/2016JG003626, 2016JG003626.
596	Kishino, M., M. Takahashi, N. Okami, and S. Ichimura (1985), Estimation of the spectral
597	absorption-coefficients of phytoplankton in the sea, Bulletin of Marine Science, 37(2),
598	634–642.
599	Lange, B. A., C. Katlein, M. Nicolaus, I. Peeken, and H. Flores (2016), Sea ice al-
600	gae chlorophyll a concentrations derived from under-ice spectral radiation profil-
601	ing platforms, Journal of Geophysical Research: Oceans, 121(12), 8511-8534, doi:
602	10.1002/2016JC011991.
603	Langhorne, P. J., K. G. Hughes, A. J. Gough, I. J. Smith, M. J. M. Williams, N. J. Robin-
604	son, C. L. Stevens, W. Rack, D. Price, G. H. Leonard, A. R. Mahoney, C. Haas, and
605	T. G. Haskell (2015), Observed platelet ice distributions in Antarctic sea ice: An index
606	for ocean-ice shelf heat flux, Geophysical Research Letters, 42(13), 5442-5451, doi:
607	10.1002/2015gl064508.
608	Legendre, L., and M. Gosselin (1991), In situ spectroradiometric estimation of microalgal
609	biomass in first-year sea ice, Polar Biology, 11(2), doi:10.1007/bf00234273.
610	Leu, E., C. Mundy, P. Assmy, K. Campbell, T. Gabrielsen, M. Gosselin, T. Juul-Pedersen,
611	and R. Gradinger (2015), Arctic spring awakening - steering principles behind the
612	phenology of vernal ice algal blooms, Progress in Oceanography, 139, 151-170, doi:
613	10.1016/j.pocean.2015.07.012.
614	McMinn, A., A. Pankowskii, C. Ashworth, R. Bhagooli, P. Ralph, and K. Ryan (2010),
615	In situ net primary productivity and photosynthesis of Antarctic sea ice algal, phy-
616	toplankton and benthic algal communities, Marine Biology, 157(6), 1345-1356, doi:
617	10.1007/s00227-010-1414-8.

618	Meiners, K. M., S. Arndt, S. Bestley, T. Krumpen, R. Ricker, M. Milnes, K. New-
619	bery, U. Freier, S. Jarman, R. King, R. Proud, S. Kawaguchi, and B. Meyer (2017),
620	Antarctic pack ice algal distribution: Floe-scale spatial variability and predictabil-
621	ity from physical parameters, Geophysical Research Letters, 44(14), 7382-7390, doi:
622	10.1002/2017gl074346.
623	Melbourne-Thomas, J., K. Meiners, C. Mundy, C. Schallenberg, K. Tattersall, and
624	G. Dieckmann (2015), Algorithms to estimate Antarctic sea ice algal biomass from
625	under-ice irradiance spectra at regional scales, Marine Ecology Progress Series, 536,
626	107-121, doi:10.3354/meps11396.
627	Melbourne-Thomas, J., K. Meiners, C. Mundy, C. Schallenberg, K. Tattersall, and
628	G. Dieckmann (2016), Corrigendum: Algorithms to estimate Antarctic sea ice algal
629	biomass from under-ice irradiance spectra at regional scales, Marine Ecology Progress
630	Series, 561, 261.
631	Mitchell, B. G. (1990), Algorithms for determining the absorption coefficient for aquatic
632	particulates using the quantitative filter technique, Proc.SPIE, 1302, 137âĂŞ-148, doi:
633	10.1117/12.21440.
634	Morecki, V. N. (1965), Underwater sea ice, Probl. Arktiki Antarktiki, 19, 32-38, (Transla-
635	tion by E.R. Hope, DRB Canada Report No. T497R April 1968).
636	Morel, A., and A. Bricaud (1981), Theoretical results concerning light absorption in a
637	discrete medium, and application to specific absorption of phytoplankton, Deep Sea Re-
638	search Part A. Oceanographic Research Papers, 28(11), 1375-1393, doi:10.1016/0198-
639	0149(81)90039-x.
640	Müller, J. L., R. R. Bidigare, C. Trees, W. M. Balch, and J. Dore (2003), Ocean Optics
641	Protocols for Satellite Ocean Colour Sensor Validation, Revision 5, Volume V: Biogeo-
642	chemical and Bio-Optical Measurements and Data, NASA Tech. Memo.
643	Mundy, C. J., D. G. Barber, and C. Michel (2005), Variability of snow and ice thermal,
644	physical and optical properties pertinent to sea ice algae biomass during spring, Journal
645	of Marine Systems, 58(3-4), 107-120, doi:10.1016/j.jmarsys.2005.07.003.
646	Mundy, C. J., J. K. Ehn, D. G. Barber, and C. Michel (2007), Influence of snow cover and
647	algae on the spectral dependence of transmitted irradiance through Arctic landfast first-
648	year sea ice, Journal of Geophysical Research, 112(C3), doi:10.1029/2006jc003683.
649	Nicolaus, M., and C. Katlein (2013), Mapping radiation transfer through sea ice using a
650	remotely operated vehicle (ROV), The Cryosphere, 7(3), 763-777, doi:10.5194/tc-7-763-

651	2013

Nicolaus, M., C. Katlein, J. Maslanik, and S. Hendricks (2012), Changes in Arctic sea ice
 result in increasing light transmittance and absorption, *Geophysical Research Letters*,
 20(24), doi:10.1020/2012gl052728

⁶⁵⁴ *39*(24), doi:10.1029/2012gl053738.

Perovich, D. K. (2017), Sea ice and sunlight, in *Sea Ice*, edited by D. N. Thomas, pp.

⁶⁵⁶ 110–137, John Wiley & Sons, Ltd, doi:10.1002/9781118778371.ch4.

- Perovich, D. K., G. F. Cota, G. A. Maykut, and T. C. Grenfell (1993), Bio-optical observations of first-year Arctic sea ice, *Geophysical Research Letters*, 20(11), 1059–1062, doi:10.1029/93gl01316.
- Smith, I. J., P. J. Langhorne, T. G. Haskell, H. J. Trodahl, R. Frew, and M. R. Vennell
 (2001), Platelet ice and the land-fast sea ice of McMurdo Sound, Antarctica, *Annals of Glaciology*, *33*(1), 21–27, doi:10.3189/172756401781818365.
- 563 Smith, I. J., A. J. Gough, P. J. Langhorne, A. R. Mahoney, G. H. Leonard, R. V. Hale,
- 664 S. Jendersie, and T. G. Haskell (2015), First-year land-fast Antarctic sea ice as an
- archive of ice shelf meltwater fluxes, *Cold Regions Science and Technology*, *113*, 63–70,
 doi:10.1016/j.coldregions.2015.01.007.
- 667 Sturm, M., and R. A. Massom (2017), Snow in the sea ice system: friend or foe?,
- in Sea Ice, edited by D. N. Thomas, pp. 65–109, John Wiley & Sons, Ltd, doi:
- 669 10.1002/9781118778371.ch3.
- Tamura, T., K. I. Ohshima, A. D. Fraser, and G. D. Williams (2016), Sea ice production
- variability in Antarctic coastal polynyas, *Journal of Geophysical Research: Oceans*,
- 672 *121*(5), 2967–2979, doi:10.1002/2015jc011537.
- ⁶⁷³ Weeks, W. (2010), *On Sea Ice*, University of Alaska Press.



Figure 1. Maps showing six transects described in this paper MS1, MS2, DS1, DS2, DS3, and DS4 with 674 12, 12, 12, 11, 10, and 10 holes, respectively. a) Scott Base in November 2014. b) Enlarged view of the 675 red patch in a). c) and d) Details of site MS1 (at 0 m: 77.668 °S; 166.524 °E, and at 512 m: 77.667 °S; 676 166.503 °E) and MS2 (at 0 m: 77.644 °S; 166.349 °E, and at 512 m: 77.646 °S; 166.331 °E), respectively, 677 in b). e) Davis Station in November 2015. f) Enlarged view of the patch in e). g) Site DS1-DS4 are 10 m 678 apart (DS1 at 0 m: 68.569 °S; 77.945 °E, and at 128 m: 68.568 °S; 77.943 °E). Note that for each transect, 679 12 holes are at 0, 0.5, 1, 2, 4, 8, 16, 32, 64, 128, 256, and 512 m except for DS3 and DS4 where transects 680 finished at 128 m. Note also that the data at 64 m DS2 were excluded from the analysis. Landsat 8 satellite 681 acquisition dates are 10 November 2014 and 17 November 2015 for Scott Base and Davis Station, respec-682 tively. Images were pan sharpened and overlaid on Quantarctica v2.0, a free GIS package for Antarctica, 683 (http://quantarctica.npolar.no/). 684



Figure 2. a) and b) Schematics of measurements in McMurdo Sound (MS1 and MS2) and Davis Station

(DS1-DS4). c) and d) Upward-looking photographs of the sea ice-water interface in McMurdo Sound and

near Davis Station (with deployed radiometer).



Figure 3. a-b) Temperature and salinity profiles of all sampling stations. c-d) and e-f) Thick and thin sec-

tions of bottom 0.1 m ice core sections from McMurdo Sound and Davis Station, respectively. Thin sections

reveal incorporated platelet ice (in d) and columnar ice structures (in f).



Figure 4. a) Plot of mean \pm 1 standard deviation of under-ice radiance spectra normalized by area under the curves in the 400 to 700 nm range with N = 24 and N = 43 for McMurdo Sound (MS) and Davis Station (DS), respectively. b) Individual under-ice radiance spectra normalized by area under the curves were plotted in the 400 to 700 nm range and grouped according to ice algal chl *a* content [mg m⁻²] in the lowermost 0.1 m of the sampled ice cores.



Figure 5. Plots of: a) a_p (the spectral absorption of all particulate matter), with standard deviation shown by shading; b) a_{nap} (the spectral absorption of non-algal particulate matter); c) a_{ph} (the spectral absorption of phytoplankton; that is $(a_p - a_{nap})$; and d) (chl *a* normalized a_{ph}^* , i.e., $a_{ph}/(chl a [mg m^{-3}])$). Note the change of scale from a) to c).



Figure 6. a) Plot of snow depth against chl *a*. b) Plot of snow depth against pigment packaging index (PPI) defined as $a_{ph}^*(440 \text{ nm})/a_{ph}^*(676 \text{ nm})$.



Figure 7. Correlation surface plots for Normalized Difference Indices (for details see text) and chl *a* and snow data. Pearson's correlation coefficient for: a) and b) combined MS and DS; c) and d) MS; and e) and f) DS for chl *a* and snow thickness. Note that values of Pearson's correlation coefficients for best wavelength pairs of combined MS and DS for chl *a* (+) and snow thickness (×) are also shown .



Figure 8. a) and b) R^2 correlation surfaces of combined MS and DS datasets for both chl *a* and snow thickness, respectively. Note that values of R^2 for best wavelength pairs for chl *a* (+) and snow thickness (×) are also shown. c) and d) Relations for best NDI(λ_1 , λ_2) against chl *a* and snow thickness, respectively. See Table 1 for the equations.



Figure 9. Comparison of empirical relationships between chl *a* and NDI without PPI (a) and with PPI (b).

Site	Ν	Empirical relationship	R^2	$^{b}R_{\rm adj}^{2}$	p-value ^c
MS	24	$\log_{10}(\text{chl }a[\text{mg m}^{-2}]) = 1.27 + 3.76 \times \text{NDI}(471,416)$	0.07	0.03	0.2
		$\text{snow}[m] = -0.20 - 1.19 \times \text{NDI}(642, 627)$	0.49	0.47	< 0.001
DS	43	$\log_{10}(\text{chl }a[\text{mg m}^{-2}]) = 2.07 - 18.16 \times \text{NDI}(439, 424)$	0.79	0.79	< 0.0001
		$\text{snow}[m] = -0.48 - 1.06 \times \text{NDI}(636, 562)$	0.70	0.69	< 0.0001
Both	67	$\log_{10}(\text{chl }a[\text{mg m}^{-2}]) = 2.58 - 16.85 \times \text{NDI}(441, 426)$	0.70	0.69	< 0.0001
		$\text{snow}[m] = -0.92 - 1.49 \times \text{NDI}(648, 567)$	0.88	0.88	< 0.0001

 Table 1.
 NDI^a summary

^{*a*}NDI stands for the normalized different index.

 ${}^{b}R_{adj}^{2} = 1 - [(N-1)/(N-P-1)](1-R^{2})$ where *P* is number of predictors excluding an intercept [e.g., *Chatterjee and Hadi*, 2012].

^{*c*} The null hypothesis of this p-value is that $\log_{10}(\operatorname{chl} a[\operatorname{mg} \mathrm{m}^{-2}]) = \operatorname{constant}$ is adequate against the empirical relationship.

Table 2. INDI-FFT Summary	Table 2.	NDI-PPI ^a	summary
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Site	Ν	Empirical relationship	R^2	$^{b}R_{\rm adj}^{2}$	p-value ^c
MS	24	$\log_{10}(\text{chl }a[\text{mg m}^{-2}]) = 0.62 + 4.40 \times \text{NDI}(471,416) + 2.24 \times \log_{10}(\text{PPI})$	0.19	0.11	0.1
DS	43	$\log_{10}(\text{chl }a[\text{mg m}^{-2}]) = 1.36 - 16.46 \times \text{NDI}(439, 424) + 2.97 \times \log_{10}(\text{PPI})$	0.81	0.80	< 0.0001
Both	67	$\log_{10}(\text{chl } a[\text{mg m}^{-2}]) = 1.44 - 14.70 \times \text{NDI}(441, 426) + 4.56 \times \log_{10}(\text{PPI})$	0.75	0.74	< 0.0001

^aNDI and PPI stand for the normalized different index, and the pigment packaging index, respectively.

 ${}^{b}R_{adj}^{2} = 1 - [(N-1)/(N-P-1)](1-R^{2})$ where P is number of predictors excluding an intercept

[e.g., Chatterjee and Hadi, 2012].

^{*c*} The null hypothesis of this p-value is that $\log_{10}(\operatorname{chl} a[\operatorname{mg} \mathrm{m}^{-2}]) = \operatorname{constant}$ is adequate against the empirical relationship.

Table 3.NDI^a comparison

Study (location)	Sea ice type	Empirical relationship	R^2	Sampling season	Mean; max chl <i>a</i> [mg m ⁻²]	Mean snow depth [m]
Present Study (McMurdo Sound, Antarctica)	fast ice (first-year)	$\log_{10}(\text{chl }a) = 1.27 + 3.76 \times \text{NDI}(471, 416)$	0.07	Austral spring	105.2; 172.4	0.01
Present Study (Davis Station, Antarctica)	fast ice (first-year)	$\log_{10}(\text{chl }a) = 2.07 - 18.16 \times \text{NDI}(439, 424)$	0.79	Austral spring	55.1; 165.0	0.27
Present Study (Both)	fast ice (first-year)	$\log_{10}(\text{chl }a) = 2.58 - 16.85 \times \text{NDI}(441, 426)$	0.70	Austral spring	73.0; 172.4	0.18
Lange et al. [2016]-Irradiance (Arctic)	pack ice (multi- and first-year)	$\ln(\text{chl}_{\text{adj}} a^b) = 2.2 + 10.8 \times \text{NDI}(683, 669)$	0.73	Arctic late summer	3.34; 11.83	< 0.1
Lange et al. [2016]-Transmittance (Arctic)	pack ice (multi- and first-year)	$\ln(\operatorname{chl}_{\operatorname{adj}} a^b) = 1.2 - 11.1 \times \operatorname{NDI}(684, 678)$	0.79	Arctic late summer	3.34; 11.83	< 0.1
Melbourne-Thomas et al. [2015] (East Antarctica, Antarctica)	pack ice (first-year)	$\ln(\text{chl } a) = -4.27 - 351 \times \text{NDI}(422, 418)$	0.64	Austral spring	0.84; 3.25	0.15
Melbourne-Thomas et al. [2015, 2016] (Weddell Sea, Antarctica)	pack ice (first-year)	$\ln(\text{chl }a) = 0.39 + 31.7 \times \text{NDI}(479, 468)$	0.79	Austral spring	6.69; 19.79	0.15
<i>Campbell et al.</i> [2014] (Allen Bay, Arctic)	fast ice (first-year)	chl $a = 15.2 - 497 \times \text{NDI}(490, 478)$	0.81	across Arctic spring	16.8; 22.1	0.16
<i>Mundy et al.</i> [2007] (Resolute Bay, Arctic)	fast ice (first-year)	chl $a = -8.3 + 1000 \times \text{NDI}(485, 472)$	0.89 ^c	Arctic spring	30.6; 109	0.16

^{*a*} NDI stands for the normalized different index. ^{*b*} chl_{adj} *a* is an adjusted chl *a*: chl_{adj} *a* = chl *a* + 0.98 mg m⁻² [*Lange et al.*, 2016]. ^{*c*} R^2 was calculated from NDI(485, 472) = 0.001 × chl *a* + 0.0083 using NDI as the predictor [*Mundy et al.*, 2007].